



**SAFETY Meeting Minutes**  
 IBC Committee  
 Zoom

**MEETING TIME RECORDS**

**Meeting start time:** 11/12/2025  
 3:00 PM  
**Meeting end time:** 4:10 PM

**VOTING MEMBER ATTENDANCE**

Name of Regular/Alternate Member	Status (Member or Alternate)	Present by Teleconference?
Karl McKinstry	Member	Yes
Gregory Danyluk	Member	Yes
Melina Kinsey	Member	Yes
Kyle Rohde	Member	Yes
Stanley Haimes	Member	Yes
Hubert Salvail	Member	Yes
Judith Hecker	Member	Yes
Lane Coffee	Member	Yes
Yulia Gerasimova	Member	Yes
Teresa Krisch	Absent	

**QUORUM INFORMATION**

**Number of SAFETY members on the roster:** 10  
**Number required for quorum:** 5

All members present by teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

**ATTENDANCE STATUS AND VOTING KEY**

<b>ABSTAIN:</b>	Present for the vote, but not voting “For” or “Against.”
<b>ABSENT:</b>	Absent for discussion and voting for reasons other than a conflicting interest.
<b>RECUSED:</b>	Absent from the meeting during discussion and voting because of a conflicting interest.
<b>SUBSTITUTION:</b>	When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member substitutes for the

	regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions.
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<b>GUEST NAMES</b>
Sophia Vermeulen, Biosafety Specialist

**Previous Meeting minutes approved:** Yes

**REVIEW OF SUBMISSIONS**

**De Novo Review**

**1. Review of SPROTO20250000014**

Title:	Doxorubicin-ES-Exosomes - Singla
Investigator:	<a href="#">Dinender Singla</a>
Submission ID	SPROTO20250000014
Funding:	• Name: National Institutes of Health (NIH), Grant Office ID: , Funding Source ID:
Agents:	• Other Cell Lines • Mouse Embryonic Stem Cells • Mouse Embryonic fibroblasts
Agent Containment:	Biological Containment Levels: • Mouse Embryonic Stem Cells: BSL-2 • Mouse Embryonic fibroblasts: BSL-2 • Other Cell Lines: BSL-2
Applicable NIH Guidelines:	NA

- a. **Description:** Doxorubicin (DOX) is an anti-cancer drug used for the treatment of various cancers. Unfortunately, use of DOX is limited due to its muscle toxicity side effects. Preliminary work and published work in our lab have shown that exosomes, specialized vesicles secreted from cells, can have beneficial effects in reducing muscle damage. These exosomes can be harvested from many cell types, including stem cells; which may provide an effective therapeutic alternative while avoiding negative side effects associated with stem cell treatment.

Exosomes will be isolated from mouse embryonic stem cells, mouse mesenchymal stem cells and mouse embryonic fibroblasts. All the cell lines were obtained from ATCC, cultured and maintained following supplier's instructions. Exosomes will be isolated using ExoQuick protocol following manufacture's instructions.

The purpose of these studies is to determine if exosomes or factors released from these exosomes can repair doxorubicin induced muscle dysfunction.

- b. **Determination:** Modifications Required

**Motion:** Lane Coffee

**Second:** Judy Hecker

**c. Required modifications:**

1. Primary Cells or Cell Lines – Please review the change made regarding the cell lines. EHS separated out the Mouse Embryonic Fibroblasts and Stem Cells from the Other Cell lines. Leaving Mouse Mesenchymal Stem Cells as an Other Cell Line.
  
2. Animals - Are animals housed in room 234 of the Biological Sciences building? If so, how long are they housed in the laboratory away from the vivarium?

**d. Votes:**

**For:** 9  
**Against:** 0  
**Recused:** 0  
**Absent:** 1  
**Abstained:** 0

**Initial Protocol**

**2. Review of SPROTO202500000016**

Title:	Bat Samples - Hoffman
Investigator:	<a href="#">Eric Hoffman</a>
Submission ID	SPROTO202500000016
Funding:	None
Agents:	• Non Human Derived Blood and Blood Types
Agent Containment:	Biological Containment Levels: • Non Human Derived Blood and Blood Types: BSL-2
Applicable NIH Guidelines:	NA

- a. **Description:** This research investigates the physiological, genetic, and ecological effects of microplastic exposure in insectivorous bats that utilize stormwater culverts and similar anthropogenic structures as roost sites in Florida. The focal species, *Tadarida brasiliensis* (Brazilian free-tailed bat), is abundant, non-listed, and commonly found in large colonies within stormwater systems, making it ideal for studying urban pollution impacts.

Bats will be captured at select stormwater sites under approved FWC permits. Biological samples will be collected non-lethally when possible (e.g., guano, oral swabs, wing punches). A subset will be humanely euthanized for tissue sampling (gut, liver, kidney, lungs, reproductive organs) to quantify internal microplastic load, conduct prey DNA analysis, and assess physiological responses, including immune

gene expression and stress biomarkers. Euthanasia will be conducted in the field, and no live bats will be brought back to the UCF Biology Department buildings.

Additional genetic screening will be conducted on Major Histocompatibility Complex (MHC) Class I and II genes to assess adaptive immunity and potential links to viral susceptibility. Salvaged carcasses of other native insectivorous bat species (*Eptesicus fuscus*, *Myotis austroriparius*, *Lasiurus intermedius*, *Nycticeius humeralis*, and *Perimyotis subflavus*) will be used for molecular comparisons when available through permitted wildlife rehabilitators. No listed species will be captured or euthanized.

- b. **Determination:** Modifications Required, Tabled, Will go back to committee after modifications

**Moved:** Karl McKinstry

**Second:** Stan Haimes

- c. **Required modifications:**

1. Summary of Research –

- a. States “Whole Specimens: Whole bat carcasses will be placed into fixative immediately in the field to ensure complete pathogen inactivation prior to transport.” How long does it take for whole bats to be fixed, and cite evidence that the time is sufficient to neutralize any pathogens? From the statement above, it states that the bats are instantly sterilized when placed in fixative.
  - b. Update the PPE section to include the following statement, “At the end of the day working in the field, ensure field clothing is removed, placed in a plastic bag until washed/bleached separately from all other clothing. The field clothing must not be brought into lab.” Or will the field PPE include disposable Tyvek suits or other disposable PPE?
  - d. Laboratory coat should be routinely laundered or use the EHS-Cintas laboratory coat exchange program. Or will you be using disposable lab coats?
  - c. Ensure handwashing is accomplished post handling bats in the field and in the laboratory. If unable to wash hands, use hand sanitizer until hands can be washed.
2. Exposure Assessment and Protective Equipment –
- a. States “All researchers on Team Bat IACUC IPROTO202500025 will have had the pre-exposure vaccines, up-to-date booster (within the last 2 years), or provide titers.” Remove “or provide titers” and replace with “and provide titers.” Personnel must have titers done at the same time

as when getting rabies booster to ensure appropriate antibody levels or personnel will be removed from the project.

- b. States. “Any employee or student that receives a bite or scratch from animals described in this document...” Edit this statement to read, “Any employee or student that receives a bite, scratch or has unprotected contact with tissues/blood to mucous membranes or compromised skin from animals described in this document...” If there is unprotected contact with tissues that have not been fixed, this should be counted as an exposure, and proper procedures need to follow.

**d. Votes:**

**For:** 9  
**Against:** 0  
**Recused:** 0  
**Absent:** 1  
**Abstained:** 0

**De Novo Review**

**3. Review of SPROTO202500000025**

Title:	Discovery and Translational Studies of Solid Nervous System Tumors - Valle
Investigator:	<a href="#">Cristina Fernandez-Valle</a>
Submission ID	SPROTO202500000025
Funding:	• Name: US Department of Defense, Grant Office ID: 2220-6A24, Funding Source ID: W81XWH2110228
Agents:	<ul style="list-style-type: none"> <li>• Human Derived Blood and Blood Types</li> <li>• Other Primary Cells</li> <li>• Lentivirus</li> <li>• Adenovirus – laboratory</li> </ul>
Agent Containment:	Biological Containment Levels: <ul style="list-style-type: none"> <li>• Other Primary Cells: BSL-2</li> <li>• Lentivirus: BSL-2</li> <li>• Adenovirus – laboratory : BSL-2</li> <li>• Human Derived Blood and Blood Types: BSL-2</li> </ul>
Applicable NIH Guidelines:	<ul style="list-style-type: none"> <li>• Section III-D-1-a</li> <li>• Section III-F-2</li> <li>• Section III-F-3</li> <li>• Section III-F-8-C-I</li> <li>• Section III-F</li> </ul>

	<ul style="list-style-type: none"> <li>• Section III-D-2-a</li> <li>• Section III-D-1</li> <li>• Section III-D-2</li> <li>• Section III-D</li> <li>• Section III-D-3</li> <li>• Section III-D-3-a</li> </ul>
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a. **Description:** The focus of the work in the Valle laboratory is to understand signalling pathways modulated by the merlin tumor suppressor to control cell proliferation and survival of schwannomas related to Neurofibromatosis. Drugs that inhibit the function of signalling proteins could be candidate therapeutics for treating Neurofibromatosis type 2 caused by loss of function of the merlin tumor suppressor. We use viral vectors to knock down gene expression to validate their role in merlin signalling pathways. We also use viral vectors to insert recombinant DNA such as to allow luciferase expression so that we can follow tumor growth in animals and measure the response to drug treatment. I conduct biomedical research using human and mouse cells and tissues.

We are conducting translational research in the lab with the goal of identifying small molecules that decrease schwannoma cell viability. We now work almost exclusively with human model schwannoma cells and patient derived schwannoma, neurofibroma and malignant peripheral nerve sheath cells and are planning to expand research to include other nervous system tumors such as glioblastoma and sarcomas. The cells are used in drug efficacy studies mostly in vitro using cell biological and biochemical assays. We have limited mouse work.

b. **Determination:** Modifications Required

Moved: Lane Coffee

Second: Hubert Salvail

c. **Required modifications:**

1. Summary of Research – States, “Normal human blood is obtained through IRB approved protocols from the Lake Nona UCF Health Center.” Please specify the clinic or hospital you are referring to by the statement of “Lake Nona UCF Health Center.”

2. Tissues, Blood or Body Fluids, Human Derived Blood and Blood Types - Source of blood is “UCF Clinical Trial Nurse.” Clarify that the blood is not from the actual clinical trial nurse and specify which patient population (demographics) samples are from.

d. **Votes:**

**For:** 9

**Against:**

**Recused:**  
**Absent:** 1  
**Abstained:**

**Initial Protocol**

**4. Review of SPROTO202500000024**

Title:	Use of human and animal blood samples for research - Huo
Investigator:	<a href="#">Qun Huo</a>
Submission ID	SPROTO202500000024
Funding:	• Name: Florida Department of Health, Grant Office ID: , Funding Source ID: MOABC
Agents:	• Human Derived Blood and Blood Types • Non Human Derived Blood and Blood Types
Agent Containment:	Biological Containment Levels: • Human Derived Blood and Blood Types: BSL-2 • Non Human Derived Blood and Blood Types: BSL-2
Applicable NIH Guidelines:	NA

- a. **Description:** We use blood samples from humans and animals to conduct biomedical research. Animal species may include livestock animals, companion animals and laboratory animal models. We develop new diagnostic tests to identify disease and health condition-related protein biomarkers. For this purpose, we need to evaluate the new tests and technology on human and animal blood samples. We also use various tests to help our research collaborators to investigate diseases-related problems using the samples we received from research collaborators. Our research involves the use of typically 10-100 microliter blood serum or plasma samples in each experiment. Our research does not involve the use of highly infectious agents or toxic biochemicals.
  
- b. **Determination:** Modifications Required  
**Moved:** Karl McKinstry  
**Second:** Stan Haimes
  
- c. **Required modifications:**
  - 1. Basic Information – Need a more specific title.
  
  - 2. Summary of Research – Clarify steps taken and where vortexing or centrifugation of blood samples are conducted to prevent the creation of aerosols.

3. Tissues, Blood, or Body Fluids – Specify the collaborators who are supplying the blood.
4. Exposure Assessment and Protective Equipment – Question 4, If there is no biosafety cabinet in laboratory, how and where do you vortex or centrifuge samples that can create aerosols? Add description of these tasks to Summary of Research.
5. Exposure Assessment and Protective Equipment – Response to potential exposure to blood, need to state that when calling Amerisys they report whether the exposure to blood was from either a Commercial (BBP clear) blood samples versus unknown (not tested) blood samples.
6. Waste Management – Question 1. Please specify the type of Lysol wipes being used to clean benchtops as some may not be sufficient for BBPs.

d. **Votes:**

<b>For:</b>	9
<b>Against:</b>	0
<b>Recused:</b>	0
<b>Absent:</b>	1
<b>Abstained:</b>	0

**REVIEW OF OTHER AGENDA ITEMS**

None