



SAFETY Meeting Minutes
 IBC Committee
 Zoom

MEETING TIME RECORDS

Meeting start time: 3/11/2026
 1:30 PM
Meeting end time: 3:02 PM

VOTING MEMBER ATTENDANCE

Name of Regular/Alternate Member	Status (Member or Alternate)	Present by Teleconference?
Karl McKinstry	Member	X
Alvina Chu	Member	Absent
Melina Kinsey	Member	X
Kyle Rohde	Member	X
Stanley Haimes	Member	X
Hubert Salvail	Member	X
Judith Hecker	Member	X
Lane Coffee	Member	Absent
Yulia Gerasimova	Member	X
Teresa Krisch	Member	X

QUORUM INFORMATION

Number of SAFETY members on the roster: 10
Number required for quorum: 5

All members present by teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

ATTENDANCE STATUS AND VOTING KEY

ABSTAIN:	Present for the vote, but not voting “For” or “Against.”
ABSENT:	Absent for discussion and voting for reasons other than a conflicting interest.
RECUSED:	Absent from the meeting during discussion and voting because of a conflicting interest.
SUBSTITUTION:	When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member substitutes for the

	regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions.
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GUEST NAMES
Sophia Vermeulen, Biosafety Specialist

Previous Meeting minutes approved: Yes

REVIEW OF SUBMISSIONS

De Novo Review

1. Review of SPROTO202600000008

Title:	Isolation of human cells from discarded surgical tissue - Kean
Investigator:	Thomas Kean
Submission ID	SPROTO202600000008
Funding:	<ul style="list-style-type: none"> • Name: National Institutes of Health (NIH), Grant Office ID: , Funding Source ID: • Name: UCF/College of Medicine, Grant Office ID: , Funding Source ID:
Agents:	<ul style="list-style-type: none"> • Skeletal Tissue • Other Primary Cells
Agent Containment:	Biological Containment Levels: <ul style="list-style-type: none"> • Other Primary Cells: BSL-2 • Skeletal Tissue: BSL-2
Applicable NIH Guidelines:	None

- a. **Description:** Discarded surgical tissue obtained from Dr. Fraser (UCF, Clinical Co-Ordinator, MTA for tissue transfer), National Disease Research Interchange (NDRI), or Gift of Hope, will be dissected and cells isolated from the tissue under sterile conditions. Typically, patients undergoing total joint replacement will have a portion of bone and cartilage removed. Our primary interest is in isolating the chondrocytes from the cartilage tissue but, as a scarce resource, we may isolate other cells from the tissue. Tissue will be received in a sterile container, in a biosafety bag, the biosafety bag is sprayed with 70% isopropanol and transferred to a biosafety cabinet. The sterile container removed from the biosafety bag and tissue retrieved with sterile forceps and transferred to a sterile plate. A scalpel is used to dissect the cartilage tissue from the joint surface and is diced into <1mm cubed pieces. These pieces are transferred to a sterile tube with media and briefly centrifuged. The supernatant is aspirated by vacuum into a container with bleach. Cells are then digested from the tissue by sequential digest with hyaluronidase and collagenase. Cells are strained through a sterile mesh (70 um) diluted and counted. These primary cells are then either plated onto cell culture plates or frozen down in 95% FBS, 5% DMSO.
- b. **Determination:** Approved with Modifications Required
Moved: Karl McKinstry
Second: Judy Hecker

c. Required modifications:

1. "Summary of Research," Need to clarify if lentivirus and reporter use is part of this BARA protocol or another BARA protocol. If they are part of this protocol, please complete the "Virus or Prions" section by adding lentivirus and complete the "Recombinant or Synthetic Nucleic Acids" section.
2. "Waste Management," Question #3: Indicates that spills in centrifuge with unsealed rotor there will be a use of respirators in this case but does not list what type of respirator that will be used. Please clarify what agents that warrant respiratory protection would be used in unsealed rotor and why buckets with biosafety seals would not be used in this situation? Additionally, are personnel part of the UCF Respiratory Protection Program? Have they been fit tested annually? Please contact EHS if need to enroll personnel in the Respiratory Protection Program.

d. Votes:

For: 8
Against: 0
Recused: 0
Absent: 2
Abstained: 0

De Novo Review

2. Review of SPROTO202600000004

Title:	Identification of human cytomegalovirus (CMV) in children's urine. - Alexander
Investigator:	Kenneth Alexander
Submission ID	SPROTO202600000004
Funding:	<ul style="list-style-type: none"> • Name: Nemours Foundation, The, Grant Office ID: , Funding Source ID: • Name: Nemours Children's Hospital, Grant Office ID: , Funding Source ID:
Agents:	<ul style="list-style-type: none"> • Cytomegalovirus (Herpesvirus) • Urine
Agent Containment:	Biological Containment Levels: <ul style="list-style-type: none"> • Cytomegalovirus (Herpesvirus): BSL-2 • Urine: BSL-2
Applicable NIH	None

Guidelines:	
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a. **Description:** In this study, we are confirming the presence of cytomegalovirus (CMV) in children known to be CMV-positive and negative in children known to be negative. We are developing a novel technique of CMV detection using the combination of non-hazardous, commercially-available immunoassays and nanotechnology, but this technology must be compared against a conventional method of CMV testing in order to validate usefulness. The conventional method utilizing quantitative real-time PCR will be used to confirm presence of CMV in the samples and will be performed in Dr. Alexander's lab. In this experimental design, with a control group, urine specimens from children in each group will be collected and processed to extract viral DNA, tested using quantitative real-time PCR to identify the presence of CMV, and then tested in our new diagnostic method. The urine is collected from children at Nemours Children's Hospital following a Nemours IRB-approved procedure.

b. **Determination:** Approved with Modifications Required

Moved: Yulia Gerasimova

Second: Karl McKinstry

c. **Required modifications:**

1. "Summary of Research" makes note of a "novel diagnostic assay." Please disclose more information about the novel assay to allow the Committee to evaluate that there are no additional biohazards.
2. "Summary of Research" indicates urine samples as being sterile, but statement is later contradicted due to the potential of CMV in urine. Best to remove the statement about urine being sterile.
3. 6 people listed in virus propagation and urine handling, but protocol team member section only has 5 members listed. This issue is a HURON issue in that the lab personnel removed from the protocol under "Protocol Team Members" are not removed from the protocol across the board. A HURON ticket has been issued for a correction. There is no action required of the PI for this finding.

d. **Votes:**

For:	8
Against:	0
Recused:	0
Absent:	2
Abstained:	0

De Novo Review

3. Review of SPROTO202600000003

Title:	Taste and Food Intake - Gilbertson
Investigator:	Timothy Gilbertson
Submission ID	SPROTO202600000003
Funding:	<ul style="list-style-type: none"> • Name: College of Medicine , Grant Office ID: DN12832, Funding Source ID: • Name: National Institutes of Health (NIH), Grant Office ID: GR105480, Funding Source ID: R43GM144020
Agents:	<ul style="list-style-type: none"> • Nervous Tissue • Other Cell Lines
Agent Containment:	Biological Containment Levels: <ul style="list-style-type: none"> • Other Cell Lines: BSL-1 • Nervous Tissue: BSL-1
Applicable NIH Guidelines:	None

a. **Description:** Our research centers on the mechanisms the body uses to recognize nutrients and how these processes play a role in determining what we choose to eat and how much we eat. Our goals are to identify the receptors for the various types of nutrients (fats, carbohydrates, proteins, and salts) and how the pathways activated by these receptors are modulated by nutritional need and by diet and disease. Using animal models we study how conditions like obesity, diabetes, and metabolic disorder change our body's ability to respond to nutrients.

b. **Determination:** Modifications Required

Moved: Teri Krisch
 Second: Karl McKinstry

c. **Required modifications:**

1. “Genetically Modified Animals: DNA Source,” The cell lines listed within Question #1 are not listed under “Primary Cells or Cell Lines.” (GPR84 KO; AdipoR1 KO; TrpM5 KO; GFP-PLCβ2; and GFP-GAD67)

d. **Votes:**

For: 8
Against: 0
Recused: 0
Absent: 2
Abstained: 0

Initial Protocol

4. Review of SPROTO20260000007

Title:	HBV detection - Gerasimova
Investigator:	Yulia Gerasimova
Submission ID	SPROTO20260000007
Funding:	• Name: National Institute of Allergy and Infectious Diseases (NIAID), Grant Office ID: , Funding Source ID:
Agents:	Recombinant DNA
Agent Containment:	BSL-2
Applicable NIH Guidelines:	• Section III-D-2-a • Section III-D-2

a. **Description:** This project aims to develop a molecular diagnostic assay for hepatitis B virus (HBV). Specifically, we will evaluate an isothermal nucleic acid amplification method, such as Nucleic Acid Sequence–Based Amplification (NASBA), for its ability to amplify a defined fragment of the HBV genome from viral DNA and/or RNA. During the assay development stage, plasmids containing HBV genomic sequences representing three major genotypes (B, C, and D), obtained from BEI Resources, will serve as templates for DNA-based amplification and for in vitro transcription to generate RNA fragments that model viral pgRNA and mRNA targets. Importantly, the plasmids used during assay optimization will not be employed for virus production or propagation in the PI’s laboratory. The resulting DNA and RNA amplification products will be characterized by gel electrophoresis and assessed using sequence-specific hybridization probes to confirm accurate target recognition across all HBV genotypes included in the study. After optimization, the assay will be evaluated using total DNA and RNA isolated from lysates of HBV-infected cells, which will be provided by Dr. Daniel Ram (UCF Burnett School of Biomedical Sciences).

b. **Determination:** Approved with Modifications Required

Moved: Stan Haimes
Second: Melina Kinsey

c. **Required modifications:**

1. “Summary of Research,” States, “After optimization, the assay will be evaluated using total DNA and RNA isolated from lysates of HBV-infected cells, which will be provided by Dr. Daniel Ram (UCF Burnett School of Biomedical Sciences).” Please clarify if the lysate collaboration with Dr. Ram is a current protocol goal or a future plan.

d. **Votes:**

For: 7
Against: 0
Recused: 1 Yulia Gerasimova
Absent: 2
Abstained: 0

Amendment

5. Review of SAMEND202600000003

Title:	Amendment for SPROTO202200000043 - Azarian
Investigator:	Taj Azarian
Submission ID	SAMEND202600000003
Applicable NIH Guidelines:	None

a. **Description:** We are amending this biosafety protocol to explicitly allow receipt and handling of human blood samples that are not in transport media. These specimens will be managed under BSL-2 practices and Universal Precautions, with all open-tube manipulations performed in a certified Class II biosafety cabinet when there is splash, splatter, or aerosol potential. The amendment adds procedures for specimen receiving and secondary containment, biohazard labeling and storage, required PPE, sharps minimization, decontamination and biomedical waste disposal, spill response, and exposure management, aligned with UCF’s Bloodborne Pathogens Exposure Control Plan.

b. **Determination:** Approved with Modifications Required

Moved: Stan Haimes
 Second: Yulia Gerasimova

c. **Required modifications:**

1. “Waste management,” Question #3, Please update with percentage and exposure time of bleach used for spill cleanup.

d. **Votes:**

For: 8
Against: 0
Recused: 0
Absent: 2
Abstained: 0

De Novo Review

6. Review of SPROTO202600000005

Title:	Borrelia burgdorferi infection mechanisms 2026-2029 - MJewett
Investigator:	Mollie Jewett
Submission ID	SPROTO202600000005
Agents:	<ul style="list-style-type: none"> • Borrelia burgdorferi • Escherichia coli K12 or derivative • Salmonella typhimurium • Human Derived Blood and Blood Types • Non Human Derived Blood and Blood Types • Human Primary Dermal Fibroblasts • J774A.1 murine-derived macrophages
Agent Containment:	<p>Biological Containment Levels:</p> <ul style="list-style-type: none"> • Escherichia coli K12 or derivative: BSL-1 • Borrelia burgdorferi: BSL-2 • Salmonella typhimurium : BSL-2 • J774A.1 murine-derived macrophages: BSL-1 • Human Primary Dermal Fibroblasts: BSL-1 • Human Derived Blood and Blood Types: BSL-2 • Non Human Derived Blood and Blood Types: BSL-2
Applicable NIH Guidelines:	<ul style="list-style-type: none"> • Section III-E-2-b • Section III-D-1-a • Section III-E-3 • Section III-D-4-b • Section III-D-4 • Section III-F • Section III-E • Section III-D-2-a • Section III-F-8-C-II • Section III-D-1 • Section III-E-2-b-(5) • Section III-D-2 • Section III-D

- a. **Description:** Borrelia burgdorferi is the bacterium that causes Lyme disease. B. burgdorferi is maintained in nature in an infectious cycle between a tick vector and small rodent hosts. B. burgdorferi is an obligate parasite and does not exist in nature outside of these hosts. Humans may acquire Lyme disease by the bite of an infected tick. We know very little about how the bacterium survives throughout its nature infectious cycle and how it causes disease in humans. The goal of B. burgdorferi infection mechanisms project in my lab is to identify bacterial components necessary for the ability of B. burgdorferi to survive during the tick-mouse infectious cycle. To do this we will use molecular genetics, immunology, biochemistry and an experimental tick-mouse infectious cycle. This research will lead to a better

- b. understanding of the biology of *B. burgdorferi* and could identify bacterial targets relevant to the diagnosis and prevention of Lyme disease.
- c. **Determination:** Approved with Modifications Required

Moved: Teri Krisch
 Second: Melina Kinsey

- d. **Required modifications:**
 1. “Summary of Research,” Need to validate that *Borrelia* can actually be completely inactivated from the FACS machine after use. Recommend the need to validate the sanitization protocol of the FACS and provide results to Chair and BSO. Contact Chair about sanitization protocol design and validation plan. Propose that there be a dedicated FACS for live infectious pathogen use.
 2. “Summary of Research,” Clarify in summary what salmonella is being used for.

- e. **Votes:**
 - For:** 8
 - Against:** 0
 - Recused:** 0
 - Absent:** 2
 - Abstained:** 0

De Novo Review

7. Review of SPROTO202500000033

Title:	Forensic Research - Ballantyne
Investigator:	John Ballantyne
Submission ID	SPROTO202500000033
Funding:	None
Agents:	<ul style="list-style-type: none"> • Semen • Vaginal Secretions • Saliva • Muscular Tissue • Human Derived Blood and Blood Types • Non Human Derived Blood and Blood Types
Agent Containment:	Biological Containment Levels: <ul style="list-style-type: none"> • Human Derived Blood and Blood Types: BSL-2 • Muscular Tissue: BSL-2 • Non Human Derived Blood and Blood Types: BSL-2 • Semen: BSL-2 • Saliva: BSL-2 • Vaginal Secretions: BSL-2

Applicable NIH Guidelines:	None
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a. **Description:** The goal of the Ballantyne/Hanson laboratory is to assist the national and international forensic science community by conducting pure and applied research in forensic molecular genetics/biochemistry in order to contribute to the body of forensic science knowledge; (2) validating methods and technologies to facilitate technology transfer, (3) providing operational support by supporting online databases of YSTR markers and mass fatality initiatives and (4) providing rigorous educational programs such as BS, MS in Forensic Science and PhDs in Chemistry and Biomolecular Science. Research areas include, but are not limited to, the development of novel Y chromosome genetic markers, tissue source identification by RNA expression profiling, the determination of individual physical characteristics by DNA typing and 'smart' single cell or low copy number analysis.

b. **Determination:** Modifications Required

Moved: Stan Haimes

Second: Judy Hecker

c. **Required modifications:**

1. "Summary of Research," extensive notes that blood and semen are coming from BioIVT and their screening process. However, there is no mention if there is screening of human blood, body fluids and skin coming from an IRB study. Need clarification if samples are screened coming from the IRB Study. How will they be handled?
2. "Tissues, Blood, or Body Fluids," Non Human Derived Blood and Blood types; Is the non-human blood from local zoos coming from sick or healthy animals. Need letter from Zoo vet that the blood is from healthy animals.
3. "Exposure Assessment and Protective Equipment," Question 1; Need to indicate that students will be notified and instructed on the potential risks of exposure to BPPs from blood and body fluids.
4. Provide notification information in the syllabus that in the case the student feels they have been exposed to a BBP they can contact the Biological Safety Officer and Student Health Services for follow up. Suggest something like the following:

For potential exposures to Bloodborne Pathogens:

- 1) Administer first aid, if necessary.
- 2) Wash the area with soap and water or flush eyes, nose or mouth with large amounts of water for 15 minutes
- 3) Notify Instructor and Biosafety Officer (407-823-1526)
- 4) Notify Student Health Services at 407-823-2701 to initiate treatment
- 5) Report the incident to EHS through this portal,

<https://live-anon.origamirisk.com/Origami/IncidentEntry/Welcome>.

d. **Votes:**

For:	8
Against:	0
Recused:	0
Absent:	2
Abstained:	0

REVIEW OF OTHER AGENDA ITEMS

None