UCF | Environmental Health and Safety
UNIVERSITY OF CENTRAL FLORIDA

Institutional Biosafety Committee Meeting Minutes
April 8, 2026
Zoom

MEETING TIME RECORDS

Meeting start time: 4/8/2026 3:00 PM
Meeting end time: 4:20 PM

VOTING MEMBER ATTENDANCE

Name of Regular/Alternate Member	Status (Member or Alternate)	Present by Teleconference?
Karl McKinstry	Member	X
Alvina Chu	Member	X
Melina Kinsey	Member	X
Kyle Rohde	Member	X
Stanley Haimes	Member	X
Hubert Salvail	Member	X
Judith Hecker	Member	Absent
Lane Coffee	Member	X
Yulia Gerasimova	Member	X
Teresa Krisch	Member	X

QUORUM INFORMATION

Number of SAFETY members on the roster: 10
Number required for quorum: 5

All members present by teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

ATTENDANCE STATUS AND VOTING KEY

ABSTAIN:	Present for the vote, but not voting "For" or "Against."
ABSENT:	Absent for discussion and voting for reasons other than a conflicting interest.
RECUSED:	Absent from the meeting during discussion and voting because of a conflicting interest.
SUBSTITUTION:	When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member substitutes for the

	regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions.

GUEST NAMES
Sophia Vermeulen, Biosafety Specialist

Previous Meeting minutes approved: Yes

February 2026 Minutes approved with minor modifications
Moved: Yulia Gerasimova
Second: Kai Karl McKinstry

March 2026 Minutes approved with minor modifications
Moved: Kai Karl McKinstry
Second: Yulia Gerasimova

REVIEW OF SUBMISSIONS

Initial Protocol

De Novo Review

1. Review of SPROTO202600000009

Title:	Immune cells for cancer immunotherapy - Copik
Investigator:	Alicja Copik
Submission ID	SPROTO202600000009
Funding:	• Name: Florida Department of Health, Grant Office ID: FP00009967, Funding Source ID: 25K07
Documents Reviewed:	• 19-27_Copik_190717.pdf • 19-27 Copik Amendment 220324.pdf • MigrationPlaceholder • MigrationPlaceholder
Agents:	• CHLA-21 • ACH2 • CHLA-136 • CEM.NKR.CCR5

	• Acinetobacter baumannii • NCI-H1385 • CHLA-52 • Escherichia coli K12 or derivative • Pseudomonas aeruginosa • CTLL-2 • COG-N-685 • Staphylococcus aureus • Nervous Tissue • Respiratory Tissue • Human Derived Blood and Blood Types • Non Human Derived Blood and Blood Types • NCI-H1915 • HL-60 • K-562 • KG-1 • KARPAS-299 • A-549 • MCF-7 • HEp-2 • L-428 • KASUMI-1 • JURKAT • HT-29 • RAJI • NCI-H810 • 22RV1 • PC-3 • NK-92 • HTB-549 • NCI-H647 • HCT-116 • L-WRN • NCI-H358 • 293T • MDA-MB-231 • NCI-H1299 • NCI-H1975 • Calu-1 • HTB-125 • HTB-126 • C4-2B • SKOV-3 • OVCAR • RWPE1 • NCI-H1703

	• NCI-H1650 • MG-63 • NCI-H1155 • BEAS-2B • Lentivirus • Adenovirus – laboratory • SK-NA-S • LAN6 • NB1643 • NB-EBc1 • SKNBE2 • CHLA-90 • BDCM • L3.6pl - WT • CHLA-255 • HFL-1 • CHLA-8 • A3.01
Agent Containment:	Biological Containment Levels: • CHLA-8: BSL-2 • NCI-H1155: BSL-2 • HTB-125: BSL-2 • NB1643: BSL-2 • L-WRN: BSL-2 • HFL-1: BSL-2 • NCI-H358: BSL-2 • CHLA-21: BSL-2 • Acinetobacter baumannii : BSL-2 • Pseudomonas aeruginosa: BSL-2 • Escherichia coli K12 or derivative: BSL-2 • 293T: BSL-2 • PC-3: BSL-2 • RAJI: BSL-2 • NK-92: BSL-2 • K-562: BSL-2 • JURKAT: BSL-2 • KARPAS-299: BSL-2 • KG-1: BSL-2 • MDA-MB-231: BSL-2 • HCT-116: BSL-2 • KASUMI-1: BSL-2 • MCF-7: BSL-2 • HL-60: BSL-2 • HT-29: BSL-2 • A-549: BSL-2 • Lentivirus: BSL-2

	• Adenovirus – laboratory : BSL-2 • C4-2B: BSL-2 • HTB-126: BSL-2 • NB-EBc1: BSL-2 • Human Derived Blood and Blood Types: BSL-2 • Respiratory Tissue: BSL-2 • Non Human Derived Blood and Blood Types: BSL-2 • Nervous Tissue: BSL-2 • NCI-H1385: BSL-2 • L3.6pl - WT: BSL-2 • SKOV-3: BSL-2 • A3.01: BSL-2 • L-428: BSL-2 • NCI-H1299: BSL-2 • SKNBE2: BSL-2 • ACH2: BSL-2 • RWPE1: BSL-2 • CHLA-52: BSL-2 • NCI-H1975: BSL-2 • OVCAR: BSL-2 • BEAS-2B: BSL-2 • CEM.NKR.CCR5: BSL-2 • CHLA-90: BSL-2 • COG-N-685 : BSL-2 • Staphylococcus aureus: BSL-2 • MG-63: BSL-2 • NCI-H1915: BSL-2 • Calu-1: BSL-2 • NCI-H1703: BSL-2 • CTLL-2: BSL-2 • CHLA-136: BSL-2 • SK-NA-S: BSL-2 • HEp-2: BSL-2 • NCI-H810: BSL-2 • NCI-H1650: BSL-2 • CHLA-255: BSL-2 • 22RV1: BSL-2 • HTB-549: BSL-2 • LAN6: BSL-2 • BDCM: BSL-2 • NCI-H647: BSL-2
Applicable NIH Guidelines:	• Section III-D-1-a • Section III-D-4-a • Section III-D-2-a • Section III-D-1 • Section III-D-2

	• Section III-D • Section III-D-3 • Section III-D-3-a

a. **Description:** Immunotherapies are revolutionizing cancer treatment. The goal of the proposed project is to design best cancer treatment strategies utilizing immune cells. This will involve use of current and development of new stimulation protocols to grow highly cytotoxic immune cells such as Natural Killer(NK) cells or gamma-delta T cells in vitro and testing their phenotype and cytotoxic function in vitro as well as their anti-cancer efficacy in vivo in murine models. Established PM21-particle and or exosome methods for NK cell stimulation will be used. Additionally new methods utilizing bacterial vesicles or membranes (e.g. from Acinetobacter baumannii and Pseudomonas aeruginosa) will be tested alone or in combination with the above established methods for the ability to enhance NK cell proliferation and cytotoxicity as well as for induction of memory-like properties in NK cells. The cells' anti-tumor function will be tested against a large variety of tumor cell lines and primary cancer cells, alone or in combination with other agents such as small molecules, nano-particles, antibodies or other biological agents. Immune cells may also be genetically (e.g. inhibitory receptor knock outs, chimeric receptor knock ins) or non genetically modified to improve their cytotoxic activity under suppressive tumor microenvironment. The antitumor activity of these modified cells will be tested in vitro in 2D and 3D spheroid models as well as in vivo in immunocompromised NSG mice seeded with human tumor cell lines or patient-derived tumor cells to develop tumors and treated with human immune cells expanded, activated and or modified in vitro. Additional treatments may be used in conjunctions such as therapeutic antibodies (e.g. anti-PD-L1), cytokines (e.g. IL2), mammalian or bacterial vesicles, ASOs etc. to improve outcomes. The methods developed for expansion of human NK cells will be also tested with monkey NK cells to be applied in the laboratory of our collaborator Dr. Ram in his research.

b. **Determination:** Modifications Required

Moved: Lane Coffee
Second: Stan Haimes

c. **Required modifications:**

1. Summary of Research: State whether the donor information (name, phone number, etc.) is available post exposure for blood samples that are not routinely screened (Advent health and UCF Health)? Or are these de-identified samples? Identifiable samples will help with treatment post exposure.

2. Primary Cells or Cell Lines: 41 Cell lines were added to the Huron database and are now noted in the Cell line list of agents. Please answer the 59 reviewer notes on these 41 cell lines so that their profiles are complete by answering questions 3, 4, 8, 10, 11 & 12 for each cell line.

d. **Votes:**

For: 9

Against: 0
Recused: 0
Absent: 1
Abstained: 0

Initial Protocol

2. Review of SPROTO202500000035

Title:	In vitro Nanoparticle Evaluation - Quadir
Investigator:	Mohi Quadir
Submission ID	SPROTO202500000035
Funding:	• Name: National Science Foundation (NSF), Grant Office ID: Not assigned yet., Funding Source ID: • Name: UCF/ENGINEERING & COMPUTER SCIENCE, COLLEGE OF (CECS), Grant Office ID: DN15182 (Start-up funding), Funding Source ID:
Documents Reviewed:	• Paper from the PI
Agents:	• Schwann Cells • HELA • Other Cell Lines • HEP-G2 • MIA-PACA-2 • MDA-MB-468 • Tumor • Blood • Panc-1 • Lentivirus • HPNE • HEK293
Agent Containment:	Biological Containment Levels: • HELA: BSL-2 • MDA-MB-468: BSL-2 • Lentivirus: BSL-2 • HPNE: BSL-2 • HEP-G2: BSL-2 • HEK293: BSL-2 • Blood: BSL-2 • Panc-1: BSL-2 • MIA-PACA-2: BSL-2 • Other Cell Lines: BSL-2 • Tumor: BSL-2 • Schwann Cells: BSL-2
Applicable NIH Guidelines:	• Section III-D-1-a • Section III-D

a. **Description:** Nanoparticles composed of polymers will be encapsulated with small molecular weight drugs, proteins, or nucleic acid. These nanoparticles will be evaluated for their particle size, surface charge, particle size distribution, and their capabilities to transport the encapsulated payload across cell membranes using physical, chemical, biochemical and microscopy-based assays.

Research Goals:

Aim 1: To test whether the nanoparticles respond to HDAC enzymes by losing acetyl groups by changing their size and structure. A) This study will help us understand whether our prepared nanoparticle is able to release the acetyl groups to downregulate HDAC activity. Our nanoparticle is designed to serve like a more potent histone by having more acetyl groups than a naturally occurring histone. This is important because histones are known as natural inhibitors of HDACs.

Aim 2: To study the behavior of HDAC specific nanoparticles. A) This aim connects the nanoparticle response to real biological conditions of the cell. This aim will also examine if the nanoparticles are safe for the cells. We will be overexpressing various HDACs in different cells which include but are not limited to HEK293 & PANC-1 and then treating them with our designed nanoparticles. This is designed to show whether the nanoparticles can sense and respond to high HDAC activity in a biological and selective way.

b. **Determination:** Approval Withheld

Moved: Kai Karl McKinstry
Second: Yulia Gerasimova

c. **Required modifications:**

1. Summary of Research: Does not align with other sections of the protocol. The research summary does not discuss the use of blood, mouse work and tumors in protocol. Sections of the summary seem to have come from a grant proposal which was not what is asked for in this section. The summary should be a short, high-level overview of the research to be performed in lay, non-technical terms. Please avoid use of acronyms and scientific jargon. Include:

- The central question the research is intended to answer
- The primary objectives
- Overview of all methods or approaches used

2. Tissues, Blood or Body Fluids:

- The tumor is marked ‘yes’ for recombinant. Is the tumor recombinant? If so, please elaborate in the “Describe the use of the agent” section.
- The Blood is marked ‘yes’ for use in animals. Will you be administering blood into animals? Or is the blood taken from animals? If the later, please mark “no” for question 10. Is agent used in animals?

3. Waste Management: Bleach treatment written as 1:13 ratio, and not 1:10 ratio. It should be a 1:10 ratio. Please correct.

d. **Votes:**

For:	9
Against:	0
Recused:	0
Absent:	1
Abstained:	0

De Novo Review

3. Review of SPROTO202600000014

Title:	Treatment of Metastatic Cancers - Khaled
Investigator:	Annette Khaled
Submission ID	SPROTO202600000014
Funding:	• Name: Florida Department of Health, Grant Office ID: AWD00006984, Funding Source ID: 25C07 • Name: Defense Health Agency (DHA), Grant Office ID: FP00009694, Funding Source ID: HT9425-25-1-0487 • Name: TBD Customer, Grant Office ID: , Funding Source ID: not applicable
Documents Reviewed:	• MigrationPlaceholder • MigrationPlaceholder • Lentiviral particles MSDS • 20-06_Khaled_200215.pdf • Salmonella • UCF Foundation
Agents:	• Baculoviruses • Salmonella typhimurium • Human Derived Blood and Blood Types • Non Human Derived Blood and Blood Types • MCF-7 • Other Cell Lines • IMR-32 • LNCAP • PC-3 • SKNAS • MDA-MB-231 • T47D • Tumor • MCF-10A • WI-38 • RWPE1 • Lentivirus
Agent Containment:	Biological Containment Levels:

	• RWPE1: BSL-2 • SKNAS: BSL-2 • WI-38: BSL-2 • T47D: BSL-2 • PC-3: BSL-2 • Baculoviruses: BSL-2 • LNCAP: BSL-2 • MCF-10A: BSL-2 • Salmonella typhimurium : BSL-2 • MCF-7: BSL-2 • MDA-MB-231: BSL-2 • IMR-32: BSL-2 • Other Cell Lines: BSL-2 • Lentivirus: BSL-2 • Non Human Derived Blood and Blood Types: BSL-2 • Human Derived Blood and Blood Types: BSL-2 • Tumor: BSL-2
Applicable NIH Guidelines:	• Section III-D-1-a • Section III-D-1 • Section III-D • Section III-D-3 • Section III-D-3-a

a. **Description:** We are researching a novel therapeutic agent in the form of a small peptide, called CT20p, to inhibit the growth of cancer cells. Death due to cancers, like breast, lung, neuroblastoma or prostate, generally results when a patient's cancer spreads from the primary site to other organs and becomes resistant to conventional treatments. This is called metastatic cancer. While current treatments for metastasis can extend the lifespan of some patients, the disease remains incurable and patients eventually die. To deliver CT20p to cancer cells, we use different delivery schemes: (1) we chemically modifiy the peptide with zwitterions, (2) we encapsulate the peptide in nanoparticles formed from a non-toxic, biodegradable polyester polymer, and (3) we bioengineer salmonella to produce the peptide tagged with EGFP. For eample, we previously showed that CT20p-nanoparticles are cytotoxic to cancer cells and cause regression of tumors in mice. Current work involves using Salmonella or similar intracellular bacteria as biological delivery vectors for CT20p or modyfing the chemistry of the peptide (e.g. zwitterions) to increased permeability. We identified the target of the CT20p peptide to be an intracellular protein called chaperonin-containing TCP-1 (CCT). We learned that the type of death caused by CT20p is immunogenic – that is activates the immune system. The goal of the research is to test whether CT20p can be targeted to tumors by forming peptide micelles, using nanoparticles or delivered using Salmonella as a biological vector, killing cancer cells while sparing normal tissue. This process also stimulates anti-cancer immunity. We will examine the specific anti-cancer effect of CT20p using immunodeficient mice that are implanted with human cancer cells and immune-enhancing activities using immunocompetent mice with syngeneic tumor cells. To track the location of the peptide micelles or nanoparticles we will use particles loaded

with non-toxic fluorescent dyes. To track Salmonella, we will use bacteria that express a tracking protein like luciferase attached to CT20p and a fluorescent marker. To optimize tumor accumulation of nanoparticles, we will coat particles with ligands targeted to receptors found on tumor cells. In parallel, we are developing the use of the CCT chaperonin as a biological indicator for cancer progression and metastasis. This involves the collect of blood or urine from cancer patients or heathy individuals and testing these biofluids for circulating tumor cells, exosomes (small vesicles shed from cells) or circulating nucleic acids that express or contain CCT protein, DNA or RNA. Fresh or fixed tissues will be used to determine CCT protein levels in tissues or exosomes as controls. The proposed research is significant because it will demonstrate the potential use of CT20p in nanoparticles or delivered by Salmonella as gene therapy as a novel cytotoxic treatment for life-threatening cancers. The development of a companion or complementary diagnostic based on detecting the CCT chaperonin can be used to monitor treatment outcomes.

b. **Determination:** Modifications Required

Moved: Melina Kinsey
Second: Stan Haimes

c. **Required modifications:**

1. Primary Cells or Cell Lines: 7 Cell lines from Question 2 were added to the Huron database and are now noted in the Cell line list of agents. Please answer the 12 reviewer notes on these 7 cell lines so that their profiles are complete by answering questions 3, 4, 8, 10, 11 & 12 for each cell line.

d. **Votes:**

For:	9
Against:	0
Recused:	0
Absent:	1
Abstained:	0

De Novo Review

4. Review of SPROTO202600000010

Title:	Blood analysis - Wells
Investigator:	Adam Wells
Submission ID	SPROTO202600000010
Funding:	None
Documents Reviewed:	• Wells MHQ.pdf • Wells BARA responses (1).docx • Previously approved BARA for the same work.
Agents:	• Human Derived Blood and Blood Types
Agent Containment:	Biological Containment Levels:

	• Human Derived Blood and Blood Types: BSL-2
Applicable NIH Guidelines:	None

a. **Description:** The resolution of muscle tissue homeostasis following exercise is mediated to a large extent by cells of the innate immune system. The research of the Cellular Exercise Physiology Laboratory (CEPL) located in BIO224 is focused on examining the cellular responses to exercise alone and in conjunction with nutritional interventions. Analyses include various whole blood, plasma and or serum constituents, including proteins, cytokines and hormones, as well as analysis of immune cell responses (surface and intracellular) through the use of flow cytometry and real time cell analysis. Characterizing the immune and inflammatory responses to different exercise modalities is important so that recovery interventions can be optimized to target specific biological outcomes that may enhance subsequent functional performance. Blood is obtained across campus in ED172 and transported to BIO 224 for storage and analysis.

b. **Determination:** Modifications Required

Moved: Kai Karl McKinstry
Second: Lane Coffee

c. **Required modifications:**

1. Short Title: Needs to be more descriptive
2. Tissues, Blood or Body Fluids: Need IRB protocol number.
3. Summary of research:

- Include the types of exercises included and examples of nutritional interventions.
- Where are specimens going to be obtained? Office? Laboratory? Clinic?
- State whether the donor information (name, phone number, etc.) is available post exposure for blood samples that are not routinely screened (IRB Protocol)? Or are these de-identified samples? Identifiable samples will help with treatment post exposure.
- Add response to reviewer comment regarding flow cytometry and blood samples to the Summary of Research. Response can be found in:
  - Tissues, Blood, or Body Fluids>Human Derived Blood>Question 3, Describe use of agent> Orange box>Reply to Dr. Rohde's comment.

d. **Votes:**

For:	9
Against:	0
Recused:	0
Absent:	1
Abstained:	0

Initial Protocol

5. Review of SPROTO202600000012

Title:	Vascular Disease-targeted Nanoparticles Evaluation - Quadir
Investigator:	Mohi Quadir
Submission ID	SPROTO202600000012
Funding:	• Name: National Science Foundation (NSF), Grant Office ID: AWD00007238, Funding Source ID: 2609681
Documents Reviewed:	None
Agents:	• Human Derived Blood and Blood Types • Other Cell Lines • HEP-G2 • 293T • HUVEC • HEK293 • HBEC WT
Agent Containment:	Biological Containment Levels: • 293T: BSL-2 • HEP-G2: BSL-2 • HUVEC: BSL-2 • HBEC WT: BSL-2 • HEK293: BSL-2 • Human Derived Blood and Blood Types: BSL-2 • Other Cell Lines: BSL-2
Applicable NIH Guidelines:	None

a. **Description:** This research is focused on creating extremely small particles made from materials that are known to be safe for the human body. The goal is to explore whether these particles could help improve future treatments for diseases that affect blood vessels. The central question of the research is whether these particles can safely interact with human blood and the cells that line blood vessels, and whether they may have characteristics that could make them useful in treating vascular diseases.

   The main objectives of the project are to design and produce these small particles and to study how safe they are when they come into contact with blood and blood clots. The research also aims to understand how the particles interact with cells that are important for the normal function of blood vessels.

   To achieve these goals, researchers will first create the particles in the laboratory using well-known methods and materials that are commonly used in biomedical studies. The particles will then be tested in carefully controlled laboratory experiments. These tests will involve growing cells that represent the type of cells found in blood vessel walls and exposing them to the particles. The particles will also be tested with samples of human blood clots.

During these experiments, the researchers will observe how the particles interact with the cells and the blood clot samples. They will evaluate whether the particles affect cell health, survival, or normal cell activity. The information gathered from these studies will help determine whether the particles appear safe and whether they may have potential for future medical applications.

b. **Determination:** Modifications Required

Moved: Yulia Gerasimova
Second: Kai Karl McKinstry

c. **Required modifications:**

1. Tissues, Blood, or Body Fluids: What is the source of the blood samples? IRB protocol or commercial source.

2. Summary of Research: Need more detail and technical information in the summary (i.e. Particle size).

d. **Votes:**

For:	9
Against:	0
Recused:	0
Absent:	1
Abstained:	0

Amendment

6. Review of SAMEND202600000002

Title:	Amendment for SPROTO202400000015 - Brown
Investigator:	Needa Brown
Submission ID	SAMEND202600000002
Funding:	None
Documents Reviewed:	None

a. **Determination:** Modifications Required

Moved: Stan Haimes
Second: Lane Coffee

b. **Required modifications:**

1. Summary of Research: Does not provide much detail on why/how the bacteria are being used. It also lists cell lines/bacteria without detailing use. The summary should be a Type a short, high-level overview of the research to be performed in lay, non-technical terms. Please avoid use of acronyms and scientific jargon. Include:

- The central question the research is intended to answer
- The primary objectives

- Overview of all methods or approaches used

2. Amendment Introduction: Question #3, Describe the rationale for the changes: The rational section does not explain why you were adding bacteria to the protocol. The Committee would also like an explanation as to why the specific drug-resistant strains are needed.

3. Primary Cells or Cell Lines: 9 Cell lines were added from Question #2 to the Huron database and are now noted in the Cell line list of agents. Please answer the 19 reviewer notes on these 9 cell lines so that their profiles are complete by answering questions 3, 4, 8, 10, 11 & 12 for each cell line.

4. Bacteria, Yeasts, Fungi, or Parasites:

- E. coli strain being used is a BSL-2 organism and needs to be listed as such and the strain is drug-resistant – Please list which drugs they are resistant to in Question #3, Describe the use of the agent.
- S. aureus strain is drug resistant – Please list which drugs they are resistant to in Question #3, Describe the use of the agent.

5. Please review and address the 24 reviewer notes within the protocol.

c. **Votes:**

For:	9
Against:	0
Recused:	0
Absent:	1
Abstained:	0

REVIEW OF OTHER AGENDA ITEMS

Melina Kinsey informed the Committee that Dr. Hoffman's bat protocol was withdrawn.